Conformational Analysis of the Cyclic Pentadepsipeptide Cyclo(Tro-Aib-Aib-Aib) in the Solid State and in Solution

by Kristian N. Koch¹), Gudrun Hopp, Anthony Linden, Kerstin Moehle, and Heinz Heimgartner*

Organisch-chemisches Institut der Universität Zürich, Winterthurerstrasse 190, CH-8057 Zürich

The cyclic 16-membered pentadepsipeptide cyclo(Tro-Aib-Aib-Aib-Aib) (1) was crystallized from MeOH/ AcOEt/CH₂Cl₂, and its structure was established by X-ray crystallography (*Fig. 1*). There are two symmetryindependent molecules with different conformations in the asymmetric unit. Two intramolecular H-bonds stabilize two β -turns in each molecule. On the other hand, two of the four Aib residues are forced to assume a nonfavorable nonhelical conformation in each of the symmetry-independent molecules (*Table 1*). The conformational study in CDCl₃ solution by NMR spectroscopy and molecular dynamics (MD) simulations indicate that the averaged structure (*Fig. 3*) is almost the same as in the solid state.

1. Introduction. – Cyclization of a peptide chain has been used extensively as a method for introducing conformational constraints in the peptide backbone. When a,a-disubstituted a-amino acids such as aminoisobutyric acid (Aib, 2,2-dimethylglycine) are incorporated into the cyclic peptide chain, the conformational flexibility of the peptide backbone is limited further.

Replacement of the H–C(α) atom by a Me group, as in the Aib residue, leads to severe restrictions of the conformational freedom [1–5]. Calculations of the allowed torsion angles ϕ and ψ for the Aib residue have shown that they are found in two very confined regions near $\phi = \pm 57^{\circ}$ and $\psi = \pm 47^{\circ}$ in the *Ramachandran* plot [3]. These two regions correspond to the right-handed and the left-handed α -helix/ β_{10} -helix, respectively. A large number of peptides containing Aib residues have had their structures established by X-ray crystallography [6–12]. It could be concluded from these structures that the Aib residue is a strong promoter of secondary structures, such as β -turns and α - and β_{10} -helices [1–5].

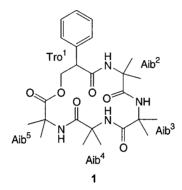
The cyclic tetrapeptide *chlamydocin* [6] and the cyclic pentapeptides cyclo(Phe-Phe-Aib-Leu-Pro) [7], cyclo(Gly-(R)-Phe(2Me)-Aib-Gly) [11], cyclo(Gly-Aib-(R)-Phe(2Me)-Aib-Gly) [12], and cyclo(Gly-(R)-Phe(2Me)-Pro-Aib-Phe) [12] each contain one Aib residue with torsion angles in the nonhelical conformational space (*i.e.*, well away from the ϕ and ψ angles mentioned above). Furthermore, Aib takes part in the formation of a γ -turn in position i+1 in some of these cyclic peptides [6][7][12], which is rather unusual for the Aib residue. These data show that Aib can be forced to assume a nonhelical conformation when this facilitates the formation of the cyclic structure and leads to the release of intramolecular strain.

Based on these precedents, it was of interest to analyze the conformation of a cyclic pentadepsipeptide containing more than one Aib residue by X-ray crystallography,

¹) Part of the Ph.D. thesis of K.N.K., Universität Zürich, 2000.

and, in solution, by NMR spectroscopy and molecular-dynamics simulations. The objective was to study the conformational preferences of the Aib residues, to determine which positions in the ϕ , ψ space they would occupy, and to compare the conformations of the cyclic depsipeptide in the solid state and in solution.

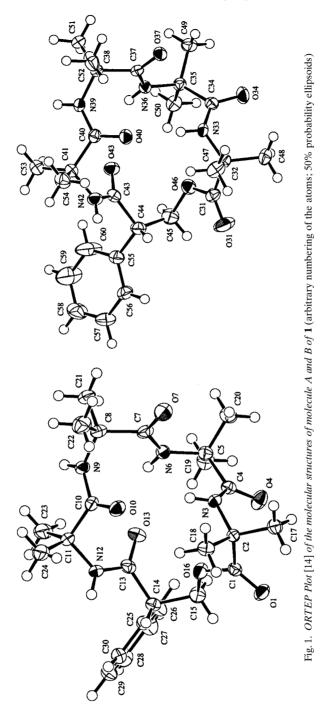
2. Results and Discussion. – The racemic cyclic pentadepsipeptide cyclo $(Tro^1-Aib^2-Aib^3-Aib^4-Aib^5)$ (**1**) was crystallized from MeOH/AcOEt/CH₂Cl₂, and its structure was established by X-ray crystallography (*Fig. 1*). The synthesis of **1** has been described previously [13].



2.1. Solid-State Conformation. There are two symmetry-independent molecules, A and B, in the asymmetric unit, and their conformations differ, both in the orientation of the Ph ring and in slightly different puckering of the macrocyclic ring. As a result of these different conformations, each of the independent molecules forms a slightly different pattern of H-bonds. The relevant torsion angles of the two molecules are given in *Table 1*; the intra- and intermolecular H-bond parameters are listed in *Table 2*.

In molecule A, the torsion angles (ϕ, ψ) for Aib³ and Aib⁵ are in good agreement with the expected values in the helical region of the *Ramachandran* plot, but the Aib² and Aib⁴ residues have torsion angles that lie in the nonhelical region. In molecule B, the torsion angles for Aib² and Aib³ were found to be similar to those of molecule A, but the torsion angles for Aib⁴ and Aib⁵ deviate significantly from the values found for molecule A: Aib⁴ has torsion angles in the helical region, whereas the torsion angles for Aib⁵ lie in the nonhelical region.

All the amide bonds have the s-*trans*-configuration ($|\omega| = 154.7 - 177.6^{\circ}$) and the ester bonds also show the s-*trans*-configuration, but some of these amide and ester bonds deviate significantly from planarity (*cf.* [15]). Each of the independent molecules A and B adopts two consecutive β -turns stabilized by the intramolecular H-bonds N(3)-H···O(10) and N(6)-H···O(13) for molecule A and N(33)-H···O(40) and N(36)-H···O(43) for molecule B (*Table 2*), although the second interaction in molecule B is extremely weak (*cf.* [16]). The combination of the torsion angles for the two residues Aib³ and Aib⁴ (ϕ_3 , ϕ_4 , and ψ_3 , ψ_4) are in good agreement with the values of a distorted β -turn conformation of type II (II) for molecule B. Molecule A forms two intermolecular H-bonds from N(9)-H and N(12)-H to O(34') and O(37''),



Residue	Angle [°]	Molecule A	Molecule B
Tro ¹	ϕ_1	156.6(2)	83.7(3)
	χ_1^{a})	46.2(3)	52.4(3)
	ψ_1	-133.6(2)	-152.5(2)
	ω_1	-176.9(2)	177.6(1)
Aib ²	ϕ_2	-55.4(3)	-52.5(3)
	ψ_2	136.3(2)	136.2(2)
	ω_2	170.2(2)	166.5(2)
Aib ³	ϕ_3	55.5(3)	51.1(3)
	ψ_3	31.4(3)	40.8(3)
	ω_3	169.2(2)	165.8(2)
Aib ⁴	ϕ_4	92.1(3)	71.0(3)
	ψ_4	-25.4(3)	23.1(3)
	ω_4	174.8(2)	154.7(2)
Aib ⁵	ϕ_5	-51.9(3)	-99.9(3)
	ψ_5	-42.9(3)	6.4(3)
	ω_5^{b})	167.3(2)	-169.5(2)

Table 1. Selected Intra-annular Torsion Angles of the Two Crystallographically Independent Molecules A and Bof 1

^a) Torsion angle of the C(2)-C(3) bond of the β -hydroxy acid Tro. ^b) Torsion angle of the lactone bond.

Molecule	Type ^a)	H…O [Å]	N…O [Å]	$N - H \cdots O[^{\circ}]$
A	$N(9)-H\cdots O(34')$	2.22(3)	3.070(3)	164(2)
	$N(12) - H \cdots O(37'')$	2.08(2)	2.957(3)	174(2)
	$N(3)-H\cdots O(10)$	2.45(2)	3.263(2)	163(2)
	$N(6) - H \cdots O(13)$	2.48(2)	3.215(3)	147(2)
В	$N(39) - H \cdots O(7'')$	2.40(2)	3.144(3)	149(2)
	$N(42) - H \cdots O(1)$	2.06(3)	2.938(3)	175(3)
	$N(33) - H \cdots O(40)$	2.33(2)	3.142(3)	162(2)
	$N(36) - H \cdots O(43)$	2.67(2)	3.367(3)	143(2)

Table 2. Selected Inter- and Intramolecular H-Bond Parameters for 1

^a) Primed atoms refer to molecules in the following symmetry-related positions: ': x, y, 1 + z; '': 1/2 - x, -1/2 + y, -1/2 - z; '': -1/2 + x, 3/2 - y, -1/2 + z.

respectively, of two different neighboring type B molecules (*Table 2*). Molecule B also forms two intermolecular H-bonds from the same amide groups, N(39)-H and N(42)-H, to two different neighboring type-A molecules, but the corresponding acceptor atoms are O(7'') and O(1), respectively. The intermolecular interactions link the molecules into a three-dimensional network (*Fig. 2*).

Recently, we reported the crystal structure of the cyclic pentadepsipeptide cyclo(Tro¹-Ac₅c²-Ac₅c³-Ac₅c⁴-Ac₅c⁵) (2) [13]. The Ac₅c residues belong to the 1-aminocycloalkane-1-carboxylic acids (Ac_nc), the family of cyclic α,α -disubstituted α -amino acids. The preferred conformations reported for the Ac_nc residues, apart from the Ac₃c residue, parallel those of the Aib residue [1][17].

In the cyclic depsipeptide 2, there are four symmetry-independent molecules in the asymmetric unit. The macrocyclic rings in the four independent molecules are very similar without significant conformational differences. The torsion angles (ϕ , ψ) for the Ac₅c residues occupying corresponding positions in each of the four independent

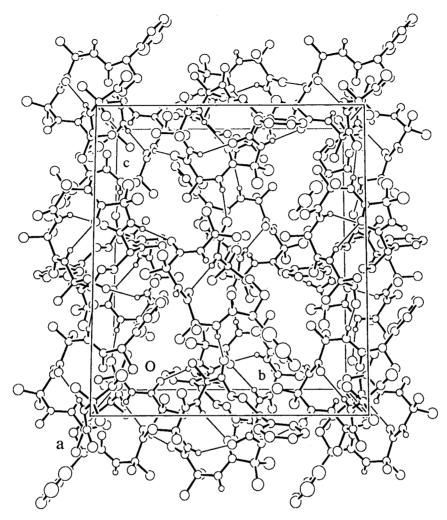
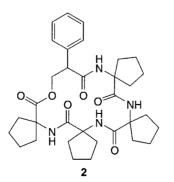


Fig. 2. Packing diagram of compound 1, showing the H-bonding



molecules are comparable. The residues Ac_5c^3 and Ac_5c^5 have a helical conformation, and those in positions 2 and 4 have a nonhelical conformation in all four independent molecules. A comparison of the torsion angles of the four Aib residues in molecule A of **1** with those of **2** reveals a pronounced similarity. Therefore, it can be concluded that the behaviors of the Aib and Ac_5c residues in the cyclic depsipeptides **1** and **2** are conformationally identical. In both cyclic depsipeptides, two of the α,α -disubstituted α amino acids have torsion angles in the nonhelical conformational space, which makes the ring formation possible and results in the release of intramolecular strain.

2.2. Solution Conformational Analysis. The conformation of the enantiomerically pure cyclic pentadepsipeptide ((R)-Tro¹-Aib²-Aib³-Aib⁴-Aib⁵) (R)-1²) in CDCl₃ solution has been studied by NMR spectroscopy. The measurements have been carried out in the temperature range of 260-300 K. In the ¹H-NMR spectrum at 275 K, only one set of sharp signals could be observed, but it could not be determined whether this was due to the presence of one major conformer, or due to a high rate of exchange between different conformers. Assignment of all the different ¹H- and ¹³C-NMR signals in (R)-1 was carried out by HSQC and HMBC techniques.

The temperature coefficients of the amide NH resonances are collected in *Table 3*, which shows the NH $\Delta \delta / \Delta T$ values measured in the range of 279-314 K. The value of NH $\Delta \delta / \Delta T$ for Aib⁴ is zero, which indicates significant shielding from the solvent. The other NH protons have values that indicate that they are more or less freely exposed to the solvent. The relative exchange rates of the amide NH protons were measured by monitoring the disappearance of their resonances upon addition of CD₃OD at 300 K. The NH proton of Aib⁴ exhibited a half-life of several minutes, whereas the other three NH protons exchanged completely within a few minutes. These two independent experiments indicate that the amide NH of Aib⁴ participates in an intramolecular H-bond, *e.g.*, the formation of a β -turn with the C=O group of Tro, which is in agreement with the solid-state structure.

	1 00	, s	, , , , , , , , , , , , , , , , , , , ,	
Residue	Aib ²	Aib ³	Aib ⁴	Aib ⁵
$-\Delta \delta / \Delta T$ [ppb/K]	4.3	- 1.1	0	1.7

Table 3. Temperature Coefficients for Amide NH of (R)-1 in the Range of 279-314 K

From the NMR ROESY experiment (T=275 K), a number of structure models of the cyclic depsipeptide (R)-1 were calculated with DYANA-1.5 [19]. This set of starting structures is rather diffuse with respect to the backbone torsion angles (without indication of the preferred solution structure), because of the missing ROE constraints of the protons at C(α). Only two significant ROE constraints (sequential distances, d_{nn}) could be used to predict the backbone orientation, namely those between NH of Aib³ and NH of Aib⁴, and between NH of Aib⁴ and NH of Aib⁵, but no other ROE constraints between non-neighboring residues were found. No further ROEs were used, because it was not possible to distinguish between the Me side chains in the Aib residue and to add corrections to the ROE distance constraints.

²) The cyclic pentadepsipeptide (*R*)-1 was prepared as previously described for racemic 1 [13] starting from (*R*)-tropic acid [18].

Beginning with the starting structure of (*R*)-1 (minimal DYANA target function), the MD simulations were performed *in vacuo* and in solution. A well-stabilized structure (*Fig. 3*) was found, which displays torsion angles (*Table 4*) that are in good agreement with those found for the two independent molecules A and B in the solid state. This structure includes two Aib-residues (Aib³, Aib⁵) in the helical conformational region. Additionally, the MD simulations indicate the presence of a strong Hbond (93%) between NH of Aib⁴ and CO of Tro¹ ($d(H \cdots O) = 2.5$ Å, N-H $\cdots O =$ 129°). This is in agreement with the experimentally found shielding of the NH of Aib⁴. The structure of conformer I is consistent with the two observed ROEs.

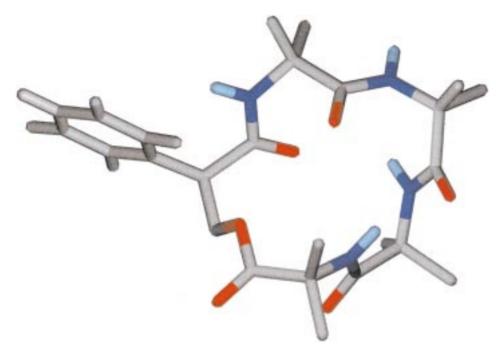


Fig. 3. Average structure obtained from MD simulations in solution

It should be mentioned that further conformations could be observed during MD simulations, starting from the variety of NMR model structures without NMR restraints. Due to the restricted flexibility of this molecule containing two methyl side chains at the $C(\alpha)$ -atoms, it is not possible to overcome more unfavored conformational regions within the simulation time scale. Further simulations were performed starting from the crystal structures A and B. Interestingly, it could be seen that both structures translate in solution into the average conformation described above. As shown in *Fig. 4*, the backbone conformations of the two crystal structures A and B are almost identical with the average solution structure, although some torsion angles for the structure B deviate significantly (*Table 4*).

3. Conclusions. – The structure and conformational behavior of the cyclic pentadepsipeptide $cyclo(Tro^1-Aib^2-Aib^3-Aib^4-Aib^5)$ (1) have been elucidated by

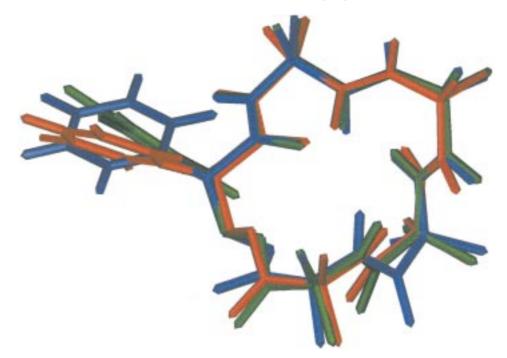


Fig. 4. Superimposition over the backbone N-, $C(\alpha)$ -, and C-atoms of the average solution structure and the crystal structures A and B (united atom model; green: crystal A; blue: crystal B; orange: average solution structure)

Table 4. Averaged Torsion Angles [°] for Conformer I of the Pentapeptide (R)-1 Obtained from MD Simulationsin Solution

Residue	ϕ	ψ	ω	χ
Tro ¹	179.9 ± 10.7	-116.7 ± 16.6	170.6 ± 7.0	54.6 ± 9.2
Aib ²	-51.9 ± 9.8	130.5 ± 16.2	-174.9 ± 6.3	
Aib ³	50.9 ± 10.7	43.8 ± 31.4	178.1 ± 7.2	
Aib ⁴	76.5 ± 11.0	-33.9 ± 10.8	179.2 ± 7.7	
Aib ⁵	-56.6 ± 10.4	-41.5 ± 12.0	$125.1 \pm 14.7^{\rm a}$)	

^a) Torsion angle of the lactone bond.

X-ray crystallography, and, in solution, by NMR spectroscopy and MD simulations. As a result of the cyclic structure, two of the four Aib residues are forced to assume a nonhelical conformation to allow release of intramolecular strain. This observation for the Aib residue is rare, because similar torsion angles (φ , ψ) for Aib have previously been observed only in the solid-state structure of cyclic tetra- and pentapeptides. A comparison of the solid-state conformation of **1** with the previously published structure of the cyclic pentadepsipeptide cyclo(Tro-Ac₅c-Ac₅c-Ac₅c-Ac₅c) (**2**) shows very similar torsion angles and conformational properties for the α , α -disubstituted α -amino acids Aib and Ac₅c. The results of the conformational analysis of **1** in CDCl₃ solution by NMR spectroscopy, followed by MD simulations, indicate that the average structure in solution at room temperature is almost the same as that in the solid state.

Experimental Part

1. X-Ray Crystal-Structure Determination of 1 (see Table 5, and Figs. 1 and 2)³). All measurements were made on a Rigaku AFC5R diffractometer with graphite-monochromated MoK_a radiation ($\lambda = 0.71069$ Å) and a 12-kW rotating anode generator. The $\omega/2\theta$ scan mode was employed for data collection. The intensities were corrected for Lorentz and polarization effects, but not for absorption. Data collection and refinement parameters are given in Table 5. A view of the molecule and a packing diagram are shown in Fig. 1 and Fig. 2, resp. The structure was solved by direct methods using SHELXS86 [20], which revealed the positions of all non-H-atoms. There are two independent molecules in the asymmetric unit, but the conformations of the molecules are significantly different, so that additional crystallographic symmetry is not possible. The non-H-atoms were refined anisotropically. All of the amide H-atoms were placed in the positions indicated by a difference electrondensity map, and their positions were fixed in geometrically calculated positions (d(C-H) = 0.95 Å), and they were assigned fixed isotropic displacement parameters with a value equal to 1.2 U_{eq} of the parent C-atom.

Table 5. Crystallographic Data of Compound 1

Crystallized from	MeOH/AcOEt/CH ₂ Cl ₂
Empirical formula	$C_{25}H_{36}N_4O_6$
Formula weight [g⋅mol ⁻¹]	488.58
Crystal color, habit	colorless, prism
Crystal dimensions [mm]	0.30 imes 0.30 imes 0.43
Temp. [K]	173(1)
Crystal system	monoclinic
Space group	$P2_1/n$
Z	8
Reflections for cell determination	25
2θ Range for cell determination [°]	23-36
Unit-cell parameters a [Å]	17.052(6)
b [Å]	16.687(5)
<i>c</i> [Å]	18.607(3)
β[°]	90.30(2)
$V[Å^3]$	5294(2)
D_x [g cm ⁻³]	1.226
$\mu(MoK_a) [mm^{-1}]$	0.0881
$2\theta_{(\text{max})}$ [°]	55
Total reflections measured	12995
Symmetry-independent reflections	12154
Reflections used $[I > 2\sigma(I)]$	7513
Parameters refined	663
Final R	0.0519
$wR (w = [\sigma(F_o) + (0.005 F_o)^2]^{-1})$	0.0451
Goodness-of-fit	1.752
Final $\Delta_{\rm max}/\sigma$	0.0003
$\Delta \rho(\max; \min) [e Å^{-3}]$	0.38; -0.25

³) Crystallographic data (excluding structure factors) for structure 1 reported here have been deposited with the *Cambridge Crystallographic Data Centre* as supplementary publication No. CCDC-145105. Copies of the data can be obtained, free of charge, on application to the CCDC, 12 Union Road, Cambridge CB21EZ, UK (fax: +44-(0)1223-336033; e-mail: deposit@ccdc.cam.ac.uk).

Refinement of the structure was carried out on F by full-matrix least-squares procedures, which minimized the function $\sum w(|F_o| - |F_c|)^2$.

Neutral-atom-scattering factors for non-H-atoms were taken from [21a], and scattering factors for H-atoms were taken from [22]. Anomalous dispersion effects were included in F_{calc} [23]; the values for f' and f'' were those of [21b]. All calculations were performed with the *TEXSAN* crystallographic software package [24].

2. NMR Measurements of (R)-1. The ROESY spectrum was measured at 275 K on a Bruker DRX-600 spectrometer by the phase-sensitive, States-TPPI method, 2D ROESY [25] experiment with cw spinlock for mixing. A data matrix of 512×2048 complex points in t_1 and t_2 , resp., were recorded. In both dimensions, data were weighted with a shifted sine-bell function and the t_1 data newly zero-filled to 1024 complex points prior to the Fourier transformation.

3. *Molecular-Dynamics Simulations of* (R)-1. Starting structures were taken from the final set of NMR structures of (R)-1. All calculations were performed with the *GROMOS96* program package in conjuction with the *GROMOS96* 43A1 force field [26]. The unusual amino acid residues Aib and Tro were created based on the similar residues Ala and Phe, respectively, with the default force field parameters and atom types.

Starting from the minimized structures, molecular-dynamics (MD) simulations were performed *in vacuo* and in solution. In case of the simulations in solution, the molecule was surrounded by CHCl₃ molecules [27] in a rectangular box (with minimum distances from the molecule to the box wall of 14 Å) under periodic boundary conditions. The MD simulations were performed by an isothermal-isobaric simulation algorithm. The temp. and the pressure were maintained (300 K and 1 atm, resp.) by weak coupling to an external temp. bath with a coupling constant of 0.1 and 0.5 ps, resp. A cut-off for the non-bonded interactions of 1.4 nm was chosen. After an equilibration period of 21 ps, the simulations were performed over a time scale of 2 ns (up to 6 ns).

REFERENCES

- [1] E. Benedetti, Biopolymers 1996, 40, 3.
- [2] C. Toniolo, M. Crisma, F. Formaggio, G. Valle, G. Cavicchioni, G. Precigoux, A. Aubry, J. Kamphuis, *Biopolymers* 1993, 33, 1061.
- [3] I. L. Karle, P. Balaram, *Biochemistry* 1990, 29, 6747.
- [4] B. V. Venkataram Prasad, P. Balaram, Crit. Rev. Biochem. 1984, 16, 307.
- [5] C. Toniolo, E. Benedetti, TIBS 1991, 16, 350.
- [6] J. L. Flippen, I. L. Karle, Biopolymers 1976, 15, 1081.
- [7] G. Zanotti, M. Saviano, G. Saviano, T. Tancredi, F. Rossi, C. Pedone, E. Benedetti, J. Pept. Res. 1998, 51, 460.
- [8] E. Escudero, X. Vidal, X. Solans, E. Peggion, J. A. Subirana, J. Pept. Sci. 1996, 2, 59.
- [9] B. Di Blasio, F. Rossi, E. Benedetti, V. Pavone, M. Saviano, C. Pedone, G. Zanotti, T. Tancredi, J. Am. Chem. Soc. 1992, 114, 8277.
- [10] F. Rossi, M. Saviano, P. Di Talia, B. Di Blasio, C. Pedone, G. Zanotti, M. Mosca, G. Saviano, T. Tancredi, K. Ziegler, E. Benedetti, *Biopolymers* 1996, 40, 465.
- [11] I. Dannecker-Dörig, Ph.D. thesis, Universität Zürich, 1995.
- [12] F. S. Arnhold, Ph.D. thesis, Universität Zürich, 1997.
- [13] K. N. Koch, A. Linden, H. Heimgartner, Helv. Chim. Acta 2000, 83, 233.
- [14] C. K. Johnson, 'ORTEPII, Report ORNL-5138', Oak Ridge National Laboratory, Oak Ridge, Tennessee, 1976.
- [15] T. Ashida, Y. Tsunogae, I. Tanaka, T. Yamane, Acta Crystallogr., Sect. B 1987, 43, 212.
- [16] R. Taylor, O. Kennard, W. Versichel, Acta Crystallogr., Sect. B 1984, 40, 280.
- [17] S. Vijavalakshmi, R. Balaji Rao, I. L. Karle, P. Balaram, Biopolymers 2000, 53, 84.
- [18] H. King, A. D. Palmer, J. Chem. Soc. 1922, 121, 2577.
- [19] P. Güntert, C. Mumenthaler, T. Herrmann, J. Mol. Biol. 1997, 273, 283.
- [20] G. M. Sheldrick, 'SHELXS86', Acta Crystallogr., Sect. A 1990, 46, 467.
- [21] a) E. N. Maslen, A. G. Fox, M. A. O'Keefe, in 'International Tables for Crystallography', Ed. A. J. C. Wilson, Kluwer Academic Publishers, Dordrecht, 1992, Vol. C, Table 6.1.1.1, pp. 477–486; b) D. C. Creagh, W. J. McAuley, *ibid*. Table 4.2.6.8, pp. 219–222.
- [22] R. F. Stewart, E. R. Davidson, W. T. Simpson, J. Chem. Phys. 1965, 42, 3175.
- [23] J. A. Ibers, W. C. Hamilton, Acta Crystallogr. 1964, 17, 781.

- [24] 'TEXSAN: Single Crystal Structure Analysis Software', Version 5.0, Molecuar Structure Corporation, The Woodlands, Texas, 1989.
- [25] A. Bax, D. G. Davis, J. Magn. Reson. 1985, 63, 207.
- [26] W. F. van Gunsteren, S. R. Billeter, A. A. Eising, P. H. Hünenberger, P. Krüger, E. A. Mark, W. R. P. Scott, I. G. Tironi, in 'Biomolecular Simulation, The GROMOS96 Manual and User Guide', Vdf Hochschulverlag AG an der ETH-Zürich, Zürich, 1996.
- [27] I. G. Tironi, W. F. van Gunsteren, Mol. Phys. 1994, 83, 381.

Received November 10, 2000